



## Variations in Germination and Seedling Traits of Rapeseed (*Brassica napus* L) Genotypes under Salinity Stress and Salicylic Acid Treatment

Jannatul Afrin<sup>1</sup>, Nikunjo Chakroborty<sup>1</sup>, Rebeka Sultana<sup>1</sup>, Khandaker Nafiz Bayazid<sup>2</sup>, Jobadatun Naher<sup>1</sup>, Arif Hasan Khan Robin<sup>1</sup>✉ 

<sup>1</sup> Department of Genetics and Plant Breeding, Bangladesh Agricultural University, Mymensingh 2202, Bangladesh

<sup>2</sup> Department of Agriculture, City University, Dhaka 1340, Bangladesh

### ARTICLE INFO

#### Article history

Received: 16 Jun 2025

Accepted: 18 Aug 2025

Published online: 25 Sep 2025

#### Keywords

Oilseed *Brassica*, Segregants, Germination phase, Trait alterations, Dual treatment, Root-shoot ratio

#### Correspondence

Arif Hasan Khan Robin

✉: [gpb21bau@bau.edu.bd](mailto:gpb21bau@bau.edu.bd)



### ABSTRACT

Rapeseed is an important oilseed crop in Bangladesh that's germination and seedling growth often get affected in saline-prone regions. This study focuses on assessing the variation in germination and seedling traits of rapeseed genotypes under salinity and dual stress – salinity with salicylic acid. Seeds of five parents and ten F<sub>3</sub> segregants of rapeseed (*Brassica napus*) were subjected to three treatments for eight days— control, 8 dSm<sup>-1</sup> salt and 8 dSm<sup>-1</sup> salt+0.1mM salicylic acid. The F<sub>3</sub> segregants were considered to evaluate the diverse inherent phenotypic expression of traits rather than only fixed genetic combination in parents. Shoot length was reduced by 52.6% and root-shoot ratio was increased by 43.7% in 8 dSm<sup>-1</sup> salinity while percent germination, shoot length, root length and root-shoot ratio were reduced by 9.6%, 53.9%, 66.7%, 27.2%, respectively, in response to 8 dSm<sup>-1</sup> salt+0.1mM salicylic acid, compared to control. Salinity stress with salicylic acid reduced percent germination, root length, dry mass per plant and root-shoot ratio by 9.1, 58.6, 20.0 and 18.3 %, respectively, compared to salinity stress alone. These results indicated that salicylic acid worsens the effects of salinity stress duration germination and early seedling growth. Despite, a few genotypes showed notable tolerance against both salinity and dual stress indicating their potential to select for further breeding and QTL analyses. PC1 explained 38% variation and separated the genotypes under 8 dSm<sup>-1</sup> salt+0.1mM salicylic acid treatment from two other treatments for high and positive coefficient for shoot length, percent germination, root length and root-shoot ratio suggesting those are salt responsive traits. The results offer a foundation for phenotypic selection, biochemical analysis, and future molecular analysis upon dual treatment of salicylic acid and salt. The results will be also useful for early identification and development of salt-resilient rapeseed lines through high resolution genetic mapping during further studies.

Copyright ©2025 by the author(s). This work is licensed under the Creative Commons Attribution International License (CC BY-NC 4.0).

## 1. Introduction

Rapeseed-mustard is the major provider of edible oil in Bangladesh as it covers the highest area of 74% of the total oilseed crops cultivated in the country (BBS 2023; Chakroborty et al. 2025). Salinity acts as a major abiotic stress in agricultural practices, which inhibits the growth of plants and reduces the yield of major crops by 50% (Bray 2000). Over 3600 million hectares of soil have been impacted by salinity, and the total area is growing by about 1.5 million hectares of land per year (Rahman et al. 2021; FAO 2024). About 37% of the total cultivable land in the southern coastal region of Bangladesh is presently affected by varying degrees of soil salinity because of tidal surges (Dasgupta et al. 2014).

Saline soil could be a potential hazard for rapeseed-mustard cultivation. While all the growth phases of a plant are impacted by salt stress; most plant species are particularly vulnerable during the seed germination and seedling growth stages (Stassinis et al. 2021; Zhang et al. 2022; BiBi et al. 2024). It was observed that seed germination percentage displayed an inverse relationship with salinity level in rapeseed-mustard (Batool et al. 2021). Failure in germination in saline soil is often associated with the accumulation of toxic ions at the seedling establishment zone for the upward movement of soil solution (imbibition) and subsequent evaporation at the soil surface (Sharma et al. 2013; Zhang et al. 2022).

### Cite This Article

Afrin J, Chakroborty N, Sultana R, Bayazid KN, Naher J, Robin AHK. 2025. Variations in Germination and Seedling Traits of Rapeseed (*Brassica napus* L) Genotypes under Salinity Stress and Salicylic Acid Treatment. *Fundamental and Applied Agriculture*, 10(3): 429–436. <https://doi.org/10.5455/faa.271230>

Two possible events e.g., either osmotic stress or ion toxicity initiated by salt stress could potentially hamper seed germination by impairing radicle growth (Sharma et al. 2013; Batool et al. 2021). The most frequent deleterious effects of salinity on *Brassica* crops include decreased plant height, growth, and yield as well as deterioration in seed quality (Ahamed et al. 2021). Salinity stress had a greater impact on the shoot & root growth, leaves, and root shoot ratio of *B. napus* (Arif et al. 2019). Salinity also induces programmed cell death (PCD) which hampers plant growth and reduces yield potential (Jalili et al. 2022).

Salicylic acid— a phenolic endogenous growth regulator, is believed to mitigate the deleterious effects of biotic and abiotic stresses in *B. napus* (Ilyas et al 2024; Raees et al 2023) while treated exogenously (seed priming or foliar spray). Salicylic acid acts as an antioxidant since it scavenges and reduces the amount of reactive oxygen species (ROS) produced in response to salt-stress (Yang et al. 2023). However, the role of salicylic acid in managing salt stress was contradictory and depended on the growth phase of plants (Afrin et al. 2025) and experimental conditions (Hayat et al. 2010). It is evident that SA regulates the root growth in a concentration dependent manner (Pour et al.; 2012, Bouallègue et al. 2017). Specific plant species and physiological stages of the plant also regulate the role of SA applied under stress (Afrin et al. 2025). Though there are several experiments which represented the role of SA in mitigating salt stress while applied through seed priming, the role of SA simultaneously applied with salt stress through the root system has not yet been understood in the germination phase of oilseed *Brassica*.

The germination rates and percentage of germinated seeds at a particular time vary considerably among species and cultivars (Mir and Somasundaram, 2021; Hasanuzzaman et al. 2010). Therefore, the treatment and genotypic variation found in rapeseed-mustard in terms of germination and early seedling traits could be exploited for introducing salt tolerant varieties (Bybordi 2010; Puppala et al.; 1999; Zheng et al. 1998).

This study investigated how salinity stress affects the germination percentage and root-shoot traits during the early germination phase in both parents and  $F_3$  segregants of *B. napus*, as well as the combining effect of salt and salicylic acid while absorbed through the root system. Despite using the single dose of SA, this study developed a baseline understanding by selecting a physiologically relevant dose of salt ( $8 \text{ dsm}^{-1}$ ) and SA ( $0.1 \text{ mM}$ ) for *B. napus* to observe the effects of dual treatments simultaneously applied in the growing media (Afrin et al. 2025). As the  $F_3$  segregants are the very early and prominent segregating populations, presence of more genetic recombination makes them potential candidate to systematically evaluate and select for quantitative traits e.g., abiotic stress resilience (Li et al. 2013). By identifying the specific morphological responses of rapeseed during germination period, the study will facilitate the early selection of suitable breeding materials from parents and  $F_3$  segregants to develop novel, salt-tolerant rapeseed genotypes.

## 2. Materials and Methods

### 2.1. Experimental site and periods with spell

The experiment was conducted in the growth room of Department of Genetics and Plant Breeding (GPB), Bangladesh Agricultural University (BAU) in Mymensingh, Bangladesh. A temperature of  $22 \pm 2^\circ \text{C}$ , relative humidity of 70%, and a 16:8-hour day: night ratio was maintained in the growth room.

### 2.2. Planting materials and source

Seeds of 10  $F_3$  generation and five parents of advanced breeding lines of rapeseed (Table 1) were collected from the Department of GPB, BAU, Mymensingh. The hybrids were the result of the previous experiments conducted at the Dept. of GPB (Azim et al. 2024).

### 2.3. Experimental design and procedure

A completely randomized design (CRD) with three treatments and three replications was adopted. There were three different treatments— control (T1),  $8 \text{ dsm}^{-1}$  salt stress (T2), and  $8 \text{ dsm}^{-1}$  salt +  $0.1 \text{ mM}$  salicylic acid (T3) (Ali et al. 2023; Göre 2025; Afrin et al. 2025). To prepare  $8 \text{ dsm}^{-1}$  salt solution, 39g of sodium chloride (NaCl) was mixed with distilled water and 0.138 mg of salicylic acid powder was added to create a concentration of  $0.1 \text{ mM}$  of salicylic acid. Moist napkin paper was used in each sterilized petri dish as growing media and ten seeds of each rapeseed genotype were placed for germination in every petri dish (Figure 1).

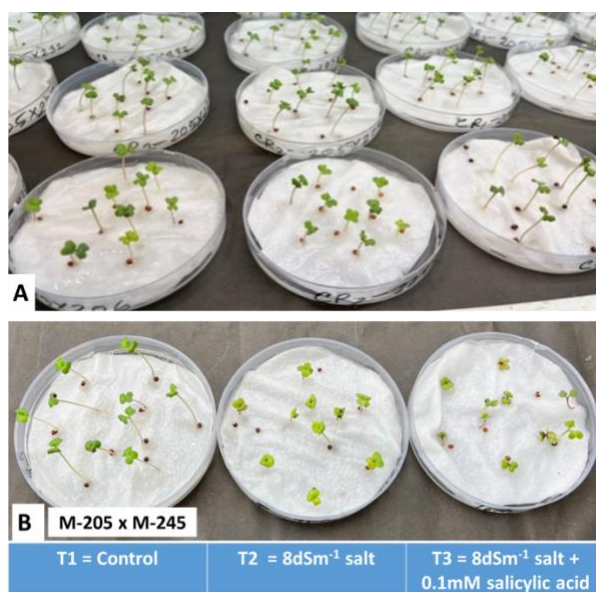


Figure 1. Germination of seeds in petri dishes in a plant culture room; (A) experimental view, (B) effect of three different level of treatments on germination and seedling growth

Table 1. Name and source of studied rapeseed genotypes

#	Code given	Genotypes	Generation	Source
1	G1	M-232×M-223	F <sub>3</sub> generation	Segregating generation developed by the research group of Professor Arif Hasan Khan Robin, Department of Genetics and Plant Breeding, Bangladesh Agricultural University
2	G2	M-205×M-232		
3	G3	M-205×M-223		
4	G4	M-223×M-206		
5	G5	M-223×M-205		
6	G6	M-205×M-245		
7	G7	M-206×M-223		
8	G8	M-232×M-245		
9	G9	M-206×M-232		
10	G10	M-245×M-206		
11	G11	M-205	Parents	Short duration, early flowering, high-yielding <i>Brassica napus</i> genotypes selected from previous experiments (Azim et al. 2024)
12	G12	M-206		
13	G13	M-223		
14	G14	M-232		
15	G15	M-245		

Treatment imposition started immediately after arranging the seeds on the petri-dishes and continued at regular intervals for eight days until destructive harvest. The majority of the seeds germinated after one or two days. The radicle of a seed was deemed to have germinated when it was at least 3 mm long. Dry weight per plant was recorded in a weighing machine after the plants were desiccated in an air-tight oven for 3 days in 60 °C.

## 2.4. Data collection and analysis

Percent germination was measured following  $(n / N) \times 100$ , where  $n$  is the number of germinated seeds at the eighth day;  $N$  is the total number of seeds. Shoot length was measured from the cotyledons to the collar region and root length was measured from the region to the root tip by using 15 cm ruler. In addition, root-shoot ratio was calculated after data collection. MINITAB 19 (Minitab Inc., State College, Pennsylvania, USA) software was used for statistical analysis. A two-way ANOVA was conducted using the General Linear Model (GLM) to examine treatment, genotype, and treatment  $\times$  genotype effects on root and shoot attributes. Tukey's pairwise comparison was used for *posthoc* analysis. Additionally, a PCA biplot was presented (Rstudio) to identify associations among genotypes and traits, and Pearson correlation analysis was performed to examine correlations between selected traits.

## 3. Results

### 3.1. Effects on germination percentage

This study explored the effects of treatment and genotype on seed germination. Both factors significantly influenced germination ( $P < 0.001$ ), though the interaction was not significant ( $P = 0.232$ ) (Table 2). The seeds under control recorded higher germination rate of 90.9% than the seeds treated with 8 dSm<sup>-1</sup> salt + 0.1 mM salicylic acid (81.3%) (Table 3). The seeds of the genotype G15 showed the highest germination rate of 98.8%, and the genotype G12 reported the lowest rate of 72.2% (Table 4).

### 3.2. Effects on Seedling Traits

The traits such as germination percentage, shoot length, root length, root-shoot ratio and dry weight plant<sup>-1</sup> were altered in the presence of salt and combined salt and

salicylic acid treatment. Analysis of variance indicated the presence of significant genotypic variation for all of the traits except for only shoot length (Table 2). Eventually, shoot length, root length and root-shoot ratio had significant against genotypic differences (Table 2). Shoot length was recorded highest 2.62 cm in control which significantly reduced when treatments were imposed i.e., 1.24 cm for 8dSm<sup>-1</sup> salt and 1.21 cm for 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (Table 3). That means shoot length was reduced significantly by 52.6% and 53.8% under 8 dSm<sup>-1</sup> salt (T2) and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (T3), respectively, compared to control (Table 2). Notably, when the impact of salicylic acid supplementation was compared with salinity stress alone it was found that 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (T3) reduced percent germination, root length, dry mass per plant and root-shoot ratio by 9.1, 58.6, 20.0 and 18.3 %, respectively, compared to salinity stress (T2) alone (Table 2).

All the genotypes except for G4 showed a clear significant reduction in shoot length in T2 and T3 compared with control (T1) (Fig. 2B). Among genotypes, significant difference was found for root length in G1, G7 within control and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (Fig. 2C). The root length of G1 was significantly reduced under 8dSm<sup>-1</sup> salt (2.57 cm) and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (0.88 cm) compared to control (6.19 cm) (Fig. 2C). The genotype G7 recorded the highest root length of 4.82 cm for control but it diminished significantly to 0.91 cm under 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (Fig. 2C). Plants under 8dSm<sup>-1</sup> salt stress recorded the highest root-shoot ratio of 2.045 and the lowest ratio of 0.837 was recorded under 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid treatment (Table 3). Among genotypes, G7 recorded the highest (1.86) root-shoot ratio and G10 and G13 had recorded the lowest (0.82) root-shoot ratio (Table 4). For the genotypes G5 and G9, the root-shoot ratio was observed to be increased by 74.5% and 71% respectively, in 8dSm<sup>-1</sup> salt stress compared to control (Fig. 2D). The genotype G5 recorded the highest root-shoot ratio of 2.99 in 8dSm<sup>-1</sup> salt than both under control (0.76 cm) and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid (0.80 cm) (Fig. 2D). Under T3, the dry weight per plant decreased significantly to 2.4 mg compared to about 4 mg in T1 and T2 (Table 3). Genotype G14 reported the highest average dry weight plant<sup>-1</sup> of 4.4 mg whereas, G5 weighed the lowest 2.2 mg (Table 4).

Table 2. Analysis of variance (mean square) for germination and seedling traits of fifteen rapeseed genotypes under salt and salt + salicylic acid induced salinity stress

Source of variation	Df	PG	SL	RL	DWP-1	R/S ratio
Genotypes (G)	14	4.87***	0.141	2.54*	0.000001***	0.89*
Treatments (T)	2	13.1***	29.16***	46.16***	0.000005***	17.6***
G × T	28	0.64	0.18**	2.04**	0.000011	0.74**
Error	90	0.52	0.091	0.93	0.000010	0.37

\*\*PG (%) =Percent germination, SL=Shoot length, RL=Root length, R/S=Root Shoot ratio, DWP<sup>-1</sup>=Dry Weight Per Plant. Different letters denote Tukey's letter of significant variation among the genotypes

Table 3. Comparison of means for germination and early seedling traits across three treatments

Traits	Control	8 dSm <sup>-1</sup> salt	8 dSm <sup>-1</sup> salt +0.1mM salicylic acid	Mean
Percent germination	90.9±0.39 <sup>a</sup>	90.4±0.26 <sup>a</sup>	81.3±0.42 <sup>b</sup>	87.53
Shoot length (cm)	2.62±0.17 <sup>a</sup>	1.24±0.11 <sup>b</sup>	1.21±0.16 <sup>b</sup>	1.69
Root length (cm)	2.95±0.69 <sup>a</sup>	2.37±0.39 <sup>a</sup>	0.98±0.17 <sup>b</sup>	2.1
Root-shoot ratio	1.15±0.28 <sup>b</sup>	2.04±0.37 <sup>a</sup>	0.837±0.17 <sup>b</sup>	1.34
Dry weight plant <sup>-1</sup> (mg)	2.98±0.00 <sup>a</sup>	2.94±0.00 <sup>a</sup>	2.4±0.00 <sup>b</sup>	2.77

Table 4. Comparison of means for germination and seedling traits in rapeseed genotypes

Traits	PG (%)	SL (cm)	RL (cm)	R/S	DWP <sup>-1</sup> (mg)
G1	87.7abc	1.64	3.2	1.7	2bc
G2	91.1a-c	1.65	2.62	1.64	3a-c
G3	86.6a-d	1.64	2.06	1.34	7a-c
G4	81.1c-e	1.65	2.15	1.34	3.3a-c
G5	84.4b-e	1.47	1.83	1.52	2.2c
G6	90a-c	1.84	1.71	0.96	3.3a-c
G7	90 a-c	1.55	2.9	1.86	2.2a-c
G8	93.3a-c	1.64	1.67	1.27	3.3a-c
G9	91.1a-c	1.55	2.22	1.67	3.3a-c
G10	95.5ab	1.71	1.3	0.82	3.3a-c
G11	90a-c	1.85	2.46	1.34	3.3a-c
G12	72.2e	1.65	1.87	1.33	3.3ab
G13	73.3de	1.88	1.38	0.82	3.3a-c
G14	87.7abc	1.73	2.21	1.25	4.4a
G15	98.8a	1.87	1.94	1.23	3.3a-c
p-value	0.001	0.998	0.215	0.285	0.001

\*\*PG (%) =Percent germination, SL=Shoot length, RL=Root length, R/S=Root Shoot ratio, DWP<sup>-1</sup>=Dry Weight Per Plant. Different letters denote Tukey's letter of significant variation among the genotypes

### 3.3. Principal component analysis

About 99% of the overall data variation for the influence of treatments and plant genotypes on five crucial germination and seedling traits were explained by the first four principal components (Table 4). PC1, PC2, PC3, and PC4 explained 41.8%, 27.5%, 16.1%, and 13.4% of the total data variation, respectively (Table 5). PC1 was highly significant for treatments, genotypes, and the treatment× genotype interaction (Table 5). For higher and positive coefficients of all the characteristics, the first principal component (PC1) distinguished rapeseed genotypes with higher germination percentage, root-shoot ratio and dry weight (Figure 5). PC1 clearly distinguished T3 (8dSm<sup>-1</sup> salt + 0.1mM salicylic acid) from other two treatments (control and 8dSm<sup>-1</sup> salt) for their lower positive coefficients for the traits e.g., percent germination, root length, shoot length and root-shoot ratio (Table 5) indicating the traits were stress-responsive (Fig. 3). Similarly, PC2 separated the genotypes under control from the genotypes treated with 8dSm<sup>-1</sup> salt and 8 dSm<sup>-1</sup>

salt + 0.1mM salicylic acid for their higher positive PC scores and higher PC coefficients of the traits stress responsive traits (Fig. 3).

### 3.4. Correlation study among germination and seedling traits

Correlation study revealed that out of ten associations, four associations were highly significant, three associations were significant, and three associations were non-significant (Fig. 4). Eight associations were positively correlated, and two associations were negatively correlated (Fig. 4). Percentage germination had highly significant ( $r>0.5$ ,  $P<0.05$ ) positive correlation with shoot length, root length and root shoot ratio (Fig. 4). Shoot length was in highly significant ( $r>0.5$ ,  $P<0.05$ ) positive correlation with root length and dry weight per plant and negative significant ( $r>0.5$ ,  $P<0.05$ ) correlation with root-shoot ratio. Root length showed highly significant ( $r>0.5$ ,  $P<0.05$ ) positive correlation with root shoot ratio (Fig. 4).



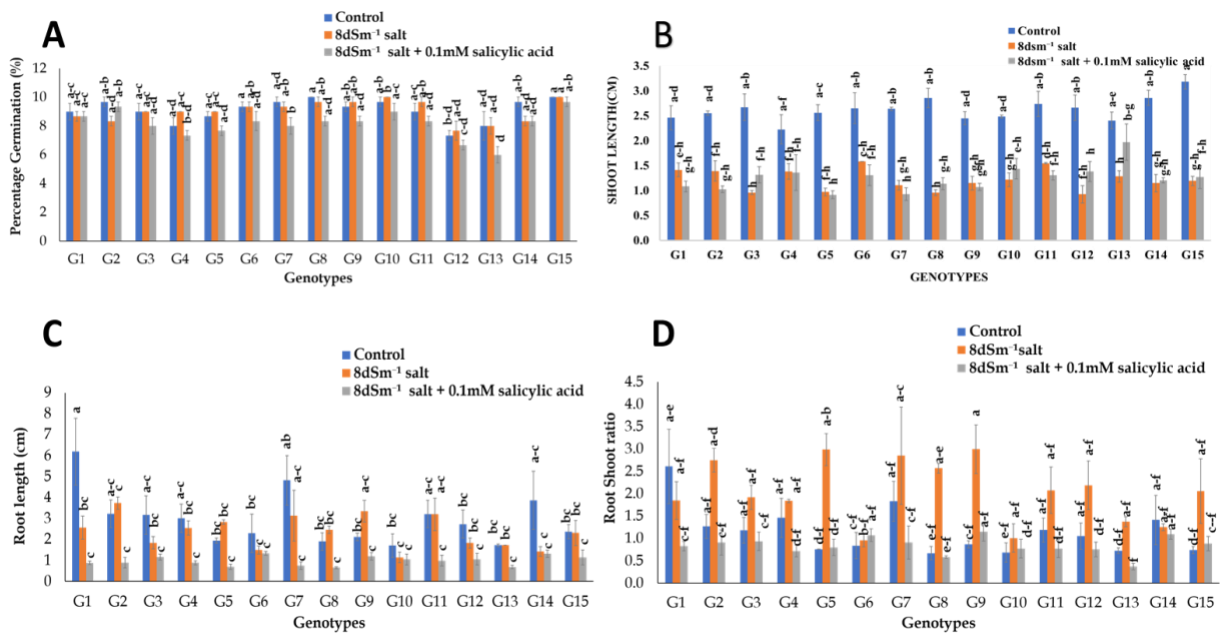


Figure 2. Treatment effect, genotypic variation and genotypic performance over environment for A) percent germination of fifteen rapeseeds genotypes under control, 8dSm<sup>-1</sup> salt and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid. B) shoot length of fifteen rapeseeds genotypes under control, 8dSm<sup>-1</sup> salt and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid. C) root length of fifteen rapeseeds genotypes under control, 8dSm<sup>-1</sup> salt and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid. D) root-shoot ratio of fifteen rapeseeds genotypes under control, 8dSm<sup>-1</sup> salt and 8dSm<sup>-1</sup> salt + 0.1mM salicylic acid. G1= M-232×M-223, G2= M-205×M-232, G3= M-205×M-223, G4= M-223×M-206, G5= M-223×M-205, G6= M-205×M-245, G7= M-206×M-223, G8= M-232×M-245, G9= M-206×M-232, G10= M-245×M-206, G11= M-205, G12= M-206, G13= M-223, G14= M-232, G15= M-245. Vertical bars indicate standard error of mean; different letters denote significant differences

Table 5. Coefficients of Principal Components for germination and seedling traits of 15 rapeseed genotypes

Variable	PC1	PC2	PC3	PC4
Percentage of Germination	0.413	0.226	-0.193	-0.860
Mean of Shoot Length	0.219	0.677	0.520	0.150
Mean of Root Length	0.676	-0.025	0.213	0.295
Dry Weight per Plant	0.076	0.519	-0.776	0.349
Root Shoot Ratio	0.564	-0.470	-0.211	0.171
Eigenvalue	1.90	1.41	0.85	0.77
% Variation explained	41.8	27.5	16.1	14.4
P-value (Genotype)	<0.001	0.075	0.258	<0.001
P-value (Treatment)	<0.001	<0.001	<0.001	0.101
P-value (Genotype× Treatment)	0.005	0.299	0.384	0.024

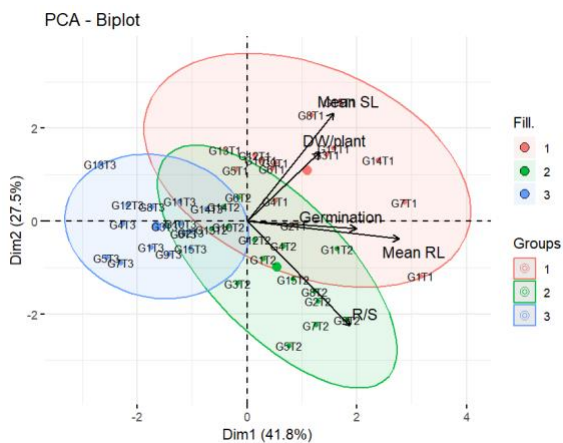


Figure 3. Biplot for germination and seedling traits of fifteen rapeseed genotypes under three treatments. G = genotype, T= Treatment, thus the combination G1T1 represent the genotype 1 under the treatment 1. Mean SL=Mean shoot length, Mean RL=Mean root length, R/S=Root Shoot ratio, DWP=1=Dry Weight Per Plant. G1= M-232×M-223, G2= M- 205×M- 232, G3= M-205×M-223, G4= M-223×M-206, G5= M-223×M-205, G6= M-205×M-245, G7= M-206×M-223, G8= M-232×M-245, G9= M-206×M-232, G10= M- 245×M-206, G11= M-205, G12= M-206, G13= M-223, G14= M-232, G15= M-245. T1= control (red), T2= 8 dSm<sup>-1</sup> salt (green) and T3= 8 dSm<sup>-1</sup> salt + 0.1mM salicylic acid (blue)

Characters	PG	SL	RL	DWP <sup>-1</sup>
SL	0.202*			
RL	0.293***	0.37***		
DWP <sup>-1</sup>	0.122 <sup>NS</sup>	0.222**	0.019 <sup>NS</sup>	
R/S	0.213**	-0.272***	0.718***	-0.077 <sup>NS</sup>

Figure 4. Heatmap showing Pearson correlation coefficients of germination and seedling traits of rapeseed genotypes. \*, \*\*, and \*\*\* indicate significance at 5%, 1%, and 0.1% probability levels, respectively. NS= non-significant. PG= Percentage of Germination, SL= Shoot Length, RL= Root Length, DWP<sup>-1</sup>= Dry Weight Per Plant, R/S= Root Shoot Ratio. Red colour represents strong positive correlation, yellow and orange colours represent weak positive correlation, green colour represent significant negative correlation which greenish colours represent non-significant correlation.

## 4. Conclusion

This study highlights the contrasting responses of *Brassica napus* genotypes to salinity and the combined application of salicylic acid (SA) with salt stress during the germination phase. While salinity alone significantly inhibited root and shoot growth, the addition of 0.1 mM SA further exacerbated stress by reducing germination percentage, root length, dry weight, and root-shoot ratio although the severity of stress was genotype-specific. Additionally, PCA identified the impact of the treatments across various rapeseed genotypes, demonstrating a strong association among the stress responsive traits through grouping (Fig. 5). PCA also showed that the genotypes G4 and G5 exhibited better performance under all treatments probably due to stress tolerance characteristics which needs further investigation.

## Acknowledgements

The primary author thanks the National Science and Technology Fellowship (2022-23) from the Ministry of Science and Technology, Government of Bangladesh. This research was supported by the Bangladesh Agricultural University Research Systems (Project no. 2021/5/BAU).

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

## References

Afrin J, Chakroborty N, Sultana R, Naher J, Robin AHK. 2025. Salicylic Acid with NaCl Acts as a Stressor and Alters Root Traits and the Estimated Root Surface Area of Rapeseed (*Brassica napus* L.) Genotypes in Hydroponic Culture. *Stresses*, 5(3), 48.  
<https://doi.org/10.3390/stresses5030048>

Ahamed F, Ahamed IM, Ahsan AS, Ahmed B, Begum F. 2021. Physiological and yield responses of some selected rapeseed/mustard genotypes to salinity stress. *Bangladesh Agronomy Journal*, 24(1): 43-55.  
<https://doi.org/10.3329/baj.v24i1.55545>

Ali E, Hussain S, Jalal F, Khan MA, Imtiaz M, Said F, Shah F. 2023. Salicylic acid-mitigates abiotic stress tolerance via altering defense mechanisms in *Brassica napus* (L.). *Frontiers in plant Science*, 14, 1187260.  
<https://doi.org/10.3389/fpls.2023.1187260>

An P, Inanaga S, Li X, Shimizu H, Tanimoto E. 2003. Root characteristics in salt tolerance. *Root Research*, 12(3): 125–132. <https://doi.org/10.3117/rootres.12.125>

Arif M, Islam M, Robin A. 2019. Salinity stress alters root morphology and root hair traits in *Brassica napus*. *Plants*, 8: 192.  
<https://doi.org/10.3390/plants8070192>

Arnon D, Hoagland D. 1944. The investigation of plant nutrition by artificial culture methods. *Biological Reviews*, 19(2): 55–67.  
<https://doi.org/10.1111/j.1469-185X.1944.tb00599.x>

Azim JB, Hassan L, Robin AHK. 2024. Genetic variation, trait association and heritability of root traits in parental and hybrid *Brassica napus* genotypes under PEG-treated hydroponic culture. Preprint. <https://doi.org/10.21203/rs.3.rs-4729831/v1>

Bahrani A, Pourreza J. 2012. Gibberellic acid and salicylic acid effects on seed germination and seedlings growth of wheat (*Triticum aestivum* L.) under salt stress condition. *World Applied Sciences Journal*, 18(5): 633–641.  
<https://doi.org/10.5829/idosi.wasj.2012.18.05.1372>

Bangladesh Bureau of Statistics (BBS). 2023. Year Book of Agricultural Statistics (2022–23). Ministry of Planning, Government of the People's Republic of Bangladesh, Dhaka, Bangladesh.

Barwal SK, Shah SH, Pawar A, Siddiqui MH, Agnihotri RK, Vimala Y, Wani SH. 2024. Mechanistic insights of salicylic acid-mediated salt stress tolerance in *Zea mays* L. seedlings. *Heliyon*, 10(14).  
<https://doi.org/10.1016/j.heliyon.2024.e34486>

Batool N, Noor T, Ilyas N, Shahzad A. 2021. Salt stress impacts on seed germination and seedling growth of *Brassica napus* L. *Pure and Applied Biology (PAB)*, 4(3): 398-406.  
<https://doi.org/10.19045/bspab.2015.43016>

BiBi R, Elahi NN, Danish S, Alahmadi TA, Ansari MJ. 2024. Enhancing germination and growth of canola (*Brassica napus* L.) through hydropriming and NaCl priming. *Scientific Reports*, 14(1): 14026. <https://doi.org/10.1038/s41598-024-63948-2>

Bouallegue A, Horchani F, Souissi F, Tebini M, Jalali K, Ahmed HB, Mhadhbi H. 2025. Enhancement of plant growth in lentil (*Lens culinaris*) under salinity stress by exogenous application or seed priming with salicylic acid and hydrogen peroxide. *PLoS One*, 20(6), e0326093.

Bouallègue A, Souissi F, Nouairi I, Souibgui M, Abbes Z, Mhadhbi H. 2017. Salicylic acid and hydrogen peroxide pretreatments alleviate salt stress in faba bean (*Vicia faba*) seeds during germination. *Seed Science and Technology*, 45: 675–690.  
<https://doi.org/10.15258/sst.2017.45.3.21>

Bray EA. 2000. Responses to abiotic stresses. In: Buchanan BB, Gruissem W, Jones RL, editors. *Biochemistry and Molecular Biology of Plants*. American Society of Plant Physiologists, Rockville, MD, USA. pp. 1158–1203.

Bybordi A. 2010. Effect of salinity and nitrogen sources on the activity of antioxidant enzymes in canola (*Brassica napus* L.). *World Applied Sciences Journal*, 8: 663–668.

Chakroborty N, Kabir MA, Hossen MM, Afrin J, Islam MM, Robin AHK. 2025. Species-Specific Variation in Oil Content and Fatty Acid Profiles of Short-Duration Rapeseed and Mustard. *ACS Food Science & Technology*, 5(3): 1082-1090.  
<https://doi.org/10.1021/acsfoodscitech.4c00894>

- Dasgupta S, Huq M, Khan ZH, Ahmed MMZ, Mukherjee N, Khan MF, Pandey K. 2014. Cyclones in a changing climate: The case of Bangladesh. *Climate and Development*, 6(2): 96–110. <https://doi.org/10.1080/17565529.2013.867242>
- Delane R, Greenway H, Munns R, Gibbs J. 1982. Ion concentration and carbohydrate status of the elongating leaf tissue of *Hordeum vulgare* growing at high external NaCl: I. Relationship between solute concentration and growth. *Journal of Experimental Botany*, 33(4): 557–573. <https://doi.org/10.1093/jxb/33.4.557>
- FAO. (2024). FAO launches first major global assessment of salt-affected soils in 50 years. 553 Newsroom. <https://www.fao.org/newsroom/detail/fao-launches-first-major-global554-assessment-of-salt-affected-soils-in-50-years/en>
- Fariduddin Q, Hayat S, Ahmad A. 2003. Salicylic acid influences net photosynthetic rate, carboxylation efficiency, nitrate reductase activity, and seed yield in *Brassica juncea*. *Photosynthetica*, 41: 281–284. <https://doi.org/10.1023/B:PHOT.0000015456.30652.c4>
- Galmés J, Molins A, Flexas J, Conesa MÀ. 2017. Coordination between leaf CO<sub>2</sub> diffusion and Rubisco properties allows maximizing photosynthetic efficiency in *Limonium* species. *Plant, Cell & Environment*, 40: 2081–2094. <https://doi.org/10.1111/pce.13000>
- Göre M. 2025. Mitigation of salt stress in *Camelina sativa* by epibrassinolide and salicylic acid treatments. *Scientific Reports*, 15(1), 7965. <https://doi.org/10.1038/s41598-025-92555-y>
- Hayat Q, Hayat S, Irfan M, Ahmad A. 2010. Effect of exogenous salicylic acid under changing environment: A review. *Environmental and Experimental Botany*, 68: 14–25. <https://doi.org/10.1016/j.envexpbot.2009.08.005>
- Hussain T, Huchzermeyer B, Koyro HW, Khan MA. 2019. Linkage between leaf development and photosynthetic response at hyperosmotic salinity in the C<sub>4</sub> grass *Panicum antidotale*. *Flora*, 256: 52–60. <https://doi.org/10.1016/j.flora.2019.02.004>
- Ilyas M, Maqsood MF, Shahbaz M, Zulfiqar U, Ahmad K, Naz N, Ali HM. 2024. Alleviating salinity stress in canola (*Brassica napus* L.) through exogenous application of salicylic acid. *BMC Plant Biology*, 24(1): 611. <https://doi.org/10.1186/s12870-024-05314-y>
- Iqbal MS, Zahoor M, Akbar M, Ahmad KS, Hussain SA, Munir S, Islam M. 2022. Alleviating the deleterious effects of salt stress on wheat (*Triticum aestivum* L.) By foliar application of gibberellic acid and salicylic acid. *Applied Ecology & Environmental Research*, 20(1). [https://doi.org/10.15666/aeer/2001\\_119134](https://doi.org/10.15666/aeer/2001_119134)
- Jalili S, Ehsanpour AA, Javadirad SM. 2022. The role of melatonin on caspase-3-like activity and expression of the genes involved in programmed cell death (PCD) induced by in vitro salt stress in alfalfa (*Medicago sativa* L.) roots. *Botanical Studies*, 63(1): 19. <https://doi.org/10.1186/s40529-022-00348-7>
- Khan MA, Ungar IA, Showalter AM. 2000. Effects of salinity on growth, water relations and ion accumulation of the subtropical perennial halophyte, *Atriplex griffithii* var. stocksii. *Annals of Botany*, 85(2): 225–232. <https://doi.org/10.1006/anbo.1999.1016>
- Kumari A, Sairam R, Singh SK, Krishna G. 2014. Early growth response: An indicator of subsequent growth and yield of wheat genotypes grown under simulated water stress condition. *Indian Journal of Plant Physiology*, 19: 94–100. <https://doi.org/10.1007/s40502-014-0088-1>
- Li L, Petsch K, Shimizu R, Liu S, Xu WW, Ying K, Muehlbauer GJ. 2013. Mendelian and non-Mendelian regulation of gene expression in maize. *PLoS genetics*, 9(1), e1003202. <https://doi.org/10.1371/journal.pgen.1003202>
- Liu J, Li L, Yuan F, Chen M. 2019. Exogenous salicylic acid improves the germination of *Limonium bicolor* seeds under salt stress. *Plant Signaling & Behavior*, 14:e1656032. <https://doi.org/10.1080/15592324.2019.1656032>
- Maas EV, Nieman RH. 1978. Physiology of plant tolerance to salinity. In: Jung GA (ed) *Crop Tolerance to Suboptimal Land Conditions*. ASA Special Publication No. 32. American Society of Agronomy, Madison, WI, USA. pp 277–299. <https://doi.org/10.2134/asapecpub32.c13>
- Michniewicz M, Zago MK, Abas L, Weijers D, Schweighofer A, Meskiene I, Heisler MG, Ohno C, Zhang J, Huang F. 2007. Antagonistic regulation of PIN phosphorylation by PP2A and PINOID directs auxin flux. *Cell*, 130, 1044–1056.
- Mir RA, Somasundaram R. 2021. Salicylic acid and salt stress tolerance in plants: a review. *Journal of Stress Physiology & Biochemistry*, 17(3):32–50.
- Munns R, Greenway H, Delane R, Gibbs J. 1982. Ion concentration and carbohydrate status of the elongating leaf tissue of *Hordeum vulgare* growing at high external NaCl: II. Cause of the growth reduction. *Journal of Experimental Botany*, 33(4):574–583. <https://doi.org/10.1093/jxb/33.4.574>
- Nonogaki H. 2010. MicroRNA gene regulation cascades during early stages of plant development. *Plant and Cell Physiology*, 51(11):1840–1846. <https://doi.org/10.1093/pcp/pcq156>
- Pasternak T, Groot EP, Kazantsev FV, Teale W, Omelyanchuk N, Kovrizhnykh V, Palme K, Mironova VV. 2019. Salicylic acid affects root meristem patterning via auxin distribution in a concentration-dependent manner. *Plant Physiology*, 180:1725–1739. <https://doi.org/10.1104/pp.19.00171>
- Pour AP, Farahbakhsh H, Saffari M, Keramat B. 2012. Effects of seed priming on germination and seedling growth under salinity stress in fenugreek. *International Journal of Agriculture and Crop Sciences*, 4:779–786.
- Puppala N, Fowler JL, Poindexter L, Bhardwaj HL. 1999. Evaluation of salinity tolerance of canola germination. In: Janick J (ed) *Perspectives on New Crops and New Uses*. ASHS Press, Alexandria, VA, USA. pp 251–253.
- Raees N, Ullah S, Nafees M. 2023. Interactive effect of tocopherol, salicylic acid and ascorbic acid on agronomic characters of two genotypes of *Brassica napus* L. under induced drought and salinity stresses. *Gesunde Pflanzen*, 75(5):1905–1923. <https://doi.org/10.1007/s10343-022-00808-x>
- Rahman MM, Mostofa MG, Keya SS, Siddiqui MN, Ansary MMU, Das AK. 2021. Adaptive mechanisms of halophytes and their potential in improving salinity tolerance in plants. *International Journal of Molecular Sciences*, 22(19):10733. <https://doi.org/10.3390/ijms221910733>
- Rajjou L, Belghazi M, Huguet R, Robin C, Moreau A, Job C, Job D. 2006. Proteomic investigation of the effect of salicylic acid on *Arabidopsis* seed germination and establishment of early defense mechanisms. *Plant Physiology*, 141:910–923. <https://doi.org/10.1104/pp.106.082057>
- Setia R, Narang S. 1985. Effects of salinity stress and plant growth regulators on seed germination and seedling growth in pea (*Pisum sativum* L.). *Flora*, 177:369–375. [https://doi.org/10.1016/S0367-2530\(17\)31621-0](https://doi.org/10.1016/S0367-2530(17)31621-0)
- Sharma P, Sardana V, Banga S. 2013. Salt tolerance of Indian mustard (*Brassica juncea*) at germination and early seedling growth. *Environmental and Experimental Biology*, 11:39–46.
- Stassinis PM, Rossi M, Borromeo I, Capo C, Beninati S, Forni C. 2021. Enhancement of *Brassica napus* tolerance to high saline conditions by seed priming. *Plants*, 10(2):403. <https://doi.org/10.3390/plants10020403>
- Waisel Y. 1985. The stimulating effects of NaCl on root growth of Rhodes grass (*Chloris gayana*). *Physiologia Plantarum*, 64:519–522. <https://doi.org/10.1111/j.1399-3054.1985.tb08532.x>
- Wang YP, Dong W, Zhang X, Yang Q, Zhang F. 2012. Effects of salicylic acid on seed germination and physiological characters of cauliflower seedlings under salt stress. In: *Proceedings of International Conference on Life Science and Technology*. pp 213–219.

- Yang W, Zhou Z, Chu Z. 2023. Emerging roles of salicylic acid in plant saline stress tolerance. *International journal of molecular sciences*, 24(4):3388. <https://doi.org/10.3390/ijms24043388>
- Zhang G, Zhou J, Peng Y, Tan Z, Li L, Yu L, Yao X. 2022. Genome-wide association studies of salt tolerance at seed germination and seedling stages in *Brassica napus*. *Frontiers in Plant Science*, 12:772708. <https://doi.org/10.3389/fpls.2021.772708>
- Zhang H, Irving LJ, McGill C, Matthew C, Zhou D, Kemp P. 2010. The effects of salinity and osmotic stress on barley germination rate: Sodium as an osmotic regulator. *Annals of Botany*, 106(6):1027–1035. <https://doi.org/10.1093/aob/mcq204>
- Zidan I, Azaizeh H, Neumann PM. 1990. Does salinity reduce growth in maize root epidermal cells by inhibiting their capacity for cell wall acidification? *Plant Physiology*, 93(1):7–11. <https://doi.org/10.1104/pp.93.1.7>